# TWO ANTHRAQUINONE GLYCOSIDES FROM CASSIA MARGINATA ROOTS

#### JANHAVI SINGH and J. SINGH

Department of Chemistry, University of Allahabad, Allahabad 211002, India

(Revised received 27 May 1986)

Key Word Index—Cassia marginata; Leguminosea; roots; anthraquinones.

Abstract—From the roots of Cassia marginata, 1,3-dihydroxy-6,8-dimethoxy-2-methyl anthraquinone 3-O-rhamnosyl- $(1 \rightarrow 6)$ -glucopyranoside and 1,3,5,8-tetrahydroxy-2-methyl anthraquinone 3-O-glucoside have been iso-lated and their structures elucidated.

#### INTRODUCTION

Species of cassia are rich sources of flavonoids [1], anthraquinones [2] and polysaccharides [3]. The plant possesses important medicinal properties [4].

# RESULTS AND DISCUSSION

Two anthraquinone glycosides (1 and 2) have been isolated from the roots of Cassia marginata Roxb.

The first glycoside, C<sub>29</sub>H<sub>34</sub>O<sub>16</sub> (1), on acid hydrolysis gave glucose, rhamnose and an aglycone, C<sub>17</sub>H<sub>14</sub>O<sub>6</sub>, which was identified as 1,3-dihydroxy-6,8-dimethoxy-2methyl anthraquinone on the basis of standard colour reactions, spectral data and co-chromatography with the natural product isolated from the seeds of C. multijuga [5]. The permethylated glycoside on hydrolysis gave two partially methylated sugars, identified as 2,3,4-tri-Omethyl glucose and 2,3,4-tri-O-methyl rhamnose by comparison of  $R_g$  values using 2,3,4,6-tetra-O-methyl-D-glucose as standard ( $R_g$  0.85 and 1.01, respectively; solvent B:A:W, 4:1:5; spray AHP). This established that two sugars, glucose and rhamnose, were present in the form of a  $1 \rightarrow 6$ -bioside [6, 7]. The  $1 \rightarrow 6$ -linkage (rutinoside) was also confirmed by the NMR spectrum of the glycoside [8, 9]. The attachment of rutinoside at position 3 was established by colour reactions and UV spectral data [10, 11]. Thus this new glycoside has been identified as 1,3-dihydroxy-6,8-dimethoxy-2-methyl anthraquinone-3-O-rhamnosyl- $(1 \rightarrow 6)$ -glucopyranoside.

The second glycoside,  $C_{21}H_{20}O_{11}$  (2), gave the characteristic colour reactions of an anthraquinone. On acid hydrolysis, it gave glucose and an aglycone,  $C_{15}H_{10}O_6$ , which on zinc dust distillation gave 2-methyl anthracene. This established that the aglycone was a 2-methyl anthraquinone derivative. The aglycone had one C-Me ( $\delta$ 2.26) and four hydroxyl groups (tetraacetate). The UV absorption maxima and mass fragments at m/z 258 [M - CO] <sup>+</sup> and 230 [M - 2CO] <sup>+</sup> were typical of an anthraquinone. The <sup>1</sup>H NMR spectrum of the aglycone showed the presence of three aromatic protons, two as an *ortho*coupled doublet at  $\delta$ 7.20 (2H, J = 9 Hz, H-6, H-7) and the third as a singlet at  $\delta$ 7.90 (1H, H-4). The aglycone was thus a 1,2,3,5,8-penta-substituted anthraquinone having a

methyl group at C-2. On the basis of standard colour reactions [12, 13], IR [14] and UV spectral data [15, 16] it was identified as 1,3,5,8-tetrahydroxy-2-methyl anthraquinone (3).

As this aglycone had not been reported earlier, its structure was confirmed by synthesis. The synthetic anthraquinone, synthesized by the cyclization of 3,6,2',4', tetrahydroxy-3'-methyl-O-benzoyl benzoic acid with sulphuric acid containing boric acid [17], was found to be identical in every respect with the aglycone isolated from the roots of C. marginata.

The attachment of the sugar moiety at position 3 was established by colour reactions and UV spectral data. The occurrence of glucose in the pyranose form was confirmed by periodate oxidation, and the  $\beta$ -nature of the sugar linkage was confirmed by  $\beta$ -glucosidase hydrolysis.

Thus the new glycoside is 1,3,5,8-tetrahydroxy-2-methyl anthraquinone  $3-O-\beta-D-(+)$ -glucopyranoside. This structure was confirmed by synthesis of the glycoside by the glycosidation of 3 in acetone in the presence of potassium hydroxide [18].

### **EXPERIMENTAL**

The roots, purchased from United Chemicals and Allied Products, Calcutta, were extracted with boiling EtOH (7 l.). The conc extract (800 ml) was poured in H<sub>2</sub>O, to give an aq. soln (fraction I) and a coloured residue (20 g fraction II).

Glycoside 2. Fraction I was concentrated (300 ml) and extracted successively with petrol, C<sub>6</sub>H<sub>6</sub>, Et<sub>2</sub>O and EtOAc. The Et<sub>2</sub>O extract (three spots on a silica gel TLC plate) was chromatographed over a silica gel column and eluted with hexane, C<sub>6</sub>H<sub>6</sub> and EtOAc. The C<sub>6</sub>H<sub>6</sub>-EtOAc (1:1) eluate (single spot on TLC) gave compound 2 (700 mg), crystallized from EtOAc-petrol (3:1), mp 190° (d). (Found: C, 56.20; H, 4.40. Calc. for C21 H20O11: C, 56.25; H, 4.46 %.) UV \( \lambda \) EtOH nm: 275, 320, 410, 480, 520; IR v<sub>max</sub> cm<sup>-1</sup>: 3450, 3272, 1610, 1455, 1360, 1230, 1080, 840, 730, 720; <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta$ 3.85 (6H, sugar protons), 5.02 (H-1' glucosyl), 2.26 (3H, s, Me), 7.88 (1H, s, H-4) and 7.20 (2H, d, J = 8.5 Hz, H-6, H-7). Acetate (Ac2O-C5H5N), mp 165°. Acid hydrolysis (7% HCl) of glycoside 2 gave an aglycone and glucose. The aglycone was crystallized from C<sub>6</sub>H<sub>6</sub>-EtOAc (4:1), mp 220°. (Found: C, 62.80; H, 3.54. Calc. for C<sub>15</sub>H<sub>10</sub>O<sub>6</sub>: C, 62.90; H, 3.49 %.) UV  $\lambda_{max}^{EIOH}$  nm: 275, 320, 410, 480, 525; IR v<sub>max</sub> cm<sup>-1</sup>: 3450, 3275, 1610, 1505, 1455, 1360, 1230, 1075; <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>): δ2.26 (s, 3H, Me), 7.90 (s, 1H, H-4), 7.20 (d, J = 8.5 Hz, H-6, H-7); MS (70 eV) m/z: 286 [M]<sup>+</sup>, 268, 258, 240, 230, 212, 183, 110, 108. Acetate (Ac2O-C5H5N), mp

Synthesis of 1,3,5,8-tetrahydroxy-2-methyl anthraquinone. 3,6-Dihydroxy phthalic anhydride (3.5 g), 2,6-dihydroxy toluene (2.4 g) and dry AlCl<sub>3</sub> (9 g) were heated at  $40-60^{\circ}$  until copious evolution of HCl gas took place. More dry AlCl<sub>3</sub> (1 g) was added and the reaction mixture was heated at  $140-160^{\circ}$  for 30 min, cooled and iced H<sub>2</sub>O (200 ml) added slowly. A dark mass was obtained, which was decomposed with cone HCl. To the residue containing benzoyl benzoic acid, a warm soln of dry Na<sub>2</sub>CO<sub>3</sub> (3 g in 50 ml H<sub>2</sub>O) was added and steam was passed into the mixture. The hot soln was filtered and the filtrate was acidified with cone HCl (5-7 ml) to liberate the acid as a brown gum.

The brown gum (3 g) was heated with conc  $H_2SO_4$  (60 ml) and boric acid (10 g) on a water bath for 80 min with occasional stirring, cooled and poured into iced  $H_2O$  (200 g). The orange-yellow solid obtained was extracted with NaHCO<sub>3</sub> soln to remove the unchanged acid, washed with  $H_2O$  and extracted with MeOH. The MeOH extract was purified by CC to give the anthraquinone (3 g), mp 220°. TLC:  $R_f$  0.75 (MeOH-EtOAc, 4:1).

Synthesis of 1,3,5,8-tetrahydroxy-2-methyl-3-O-glucopyranoside. To a soln of the above-synthesized anthraquinone (200 mg) and acetobromoglucose (600 mg) in Me<sub>2</sub>CO (50 ml), a soln of KOH (150 mg) in H<sub>2</sub>O (17 ml) was added dropwise. After stirring for 2 hr, the Me<sub>2</sub>CO was evaporated and the residue extracted with CHCl<sub>3</sub>. The CHCl<sub>3</sub> soln was washed with H<sub>2</sub>O, dried and the solvent evaporated. The acetylated product in MeOH (10 ml) was treated with M NaOMe (10.5 ml) at room temp. for 24 hr and subsequently acidified (HOAc) to pH 6. The product was purified by CC to yield light-brown crystals from HOAc, 120 mg (28 %), mp 188° (lit. 190°).

Glycoside 1. The EtOAc extract of the roots of C. marginata was chromatographed over silica gel and eluted successively with

hexane, C<sub>6</sub>H<sub>6</sub> and EtOAc. The C<sub>6</sub>H<sub>6</sub>-EtOAc eluate (single spot on TLC) gave a vellow-coloured compound (1), 800 mg, crystallized from EtOAc-petrol (1:5), mp 120° (d). (Found: C, 47.08; H, 4.70. Calc. for  $C_{29}H_{34}O_{16}$ : C, 47.15; H, 4.61%.) UV  $\lambda_{max}^{ErOH}$  nm: 222, 250, 373, 440; IR  $\nu_{max}^{KBr}$  cm<sup>-1</sup>: 3340, 2945, 2892, 1765, 1725, 1650, 1185, 730, 720; <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>): δ0.98 (br, rhamnose methyl), 2.10 (s, Me), 3.85 (s, 6H-OMe), 3.40-3.85 (br, sugar protons), 5.00 (H-l" rhamnosyl), 5.25 (H-l" glucosyl), 6.85 (d, J = 2.5 Hz, 1H, H-7), 7.68 (d, J = 2.5 Hz, 1H, H-5), 7.85 (s, 1H, H-4). Acetate (Ac<sub>2</sub>O-C<sub>5</sub>H<sub>5</sub>N) mp 260°. Permethylation of 1 was carried out by the method described in ref. [19]. The characterization of the permethylated sugars is described in the Results and Discussion. Acid hydrolysis of glycoside 1 (400 mg) with 7% HCl (3 ml) was performed for 4 hr. The mixture was then diluted with H<sub>2</sub>O and the soln extracted with Et<sub>2</sub>O (100 ml). The aq. layer was chromatographed on Whatman No. 1 paper in (a) EtOAc-C<sub>5</sub>H<sub>5</sub>N-H<sub>2</sub>O (12:5:4) and (b) EtOAc-i-PrOH-H<sub>2</sub>O (3:1:1) using glucose, rhamnose, arabinose, and xylose as standards. The chromatogram was developed with p-anisidine HCl (1 g) and NaHS (0.1 g) in MeOH (10 ml) diluted to 100 ml with n-BuOH and aniline hydrogen phthalate spray at 120-130° for 10-15 min. The chromatogram showed the presence of glucose and rhamnose ( $R_f$  0.18, 0.34). The Et<sub>2</sub>O extract was evaporated to yield the aglycone, crystallized from EtOAc-petrol (3:1) mp 272°. UV  $\lambda_{\rm max}^{\rm EiOH}$  nm: 220, 250, 285, 375, 440; MS (70 eV) m/z: 314 [M]+, 296, 286, 268, 258, 240, 229, 136 and 122. Acetate  $(Ac_2O-C_5H_5N)$ , mp 230°.

Acknowledgements—One of us (J.S.) is grateful to the C.S.I.R., India for the award of a Junior Research Fellowship, to Dr. P. D. Satsangi, and to the Pennsylvania State University, U.S.A., for mass and NMR spectra.

# REFERENCES

- 1. Tiwari, R. D. and Singh, J. (1978) Planta Med. 34, 319.
- 2. Tiwari, R. D. and Singh, J. (1978) Phytochemistry 18, 347.
- 3. Gupta, P. C. and Dubey, P. (1978) Carbohydr. Res. 72, 151.
- Chopra, R. N., Chopra, I. C., Hande, K. L. and Kapur, L. D. (1958) Indigenous Drugs of India, pp. 98, 499. Dhar.
- 5. Singh, J. (1981) Planta Med. 41, 397.
- Horhammer, L. and Hansel, R. (1955) Arch. Pharm. Berlin 288, 315.
- Hirst, E. L. and Jones, J. K. N. (1949) Discuss. Faraday Soc. 7, 268.
- Kutney, J. P., Warnock, W. D. C. and Gilbert, B. (1970) *Phytochemistry* 9, 1877.
- Sherwood, R. T. and Sharma, M. (1973) Phytochemistry 12, 2275.
- 10. Graebe, C. (1906) Annalen 211, 349.
- Shibata, S., Takido, M. and Tanaka, O. (1950) J. Am. Chem. Soc. 72, 2789.
- 12. Lemli, J. and Sequeker, R. (1964) J. Pharm. Weakblad 49, 357.
- Briggs, L. H., Jacombs, F. B. and Nicholls, G. A. (1953) J. Chem. Soc. 3069.
- Thomson, R. H. (1971) Naturally Occurring Quinones, 2nd edn. Academic Press, London.
- Briggs, L. H., Nicholls, G. A. and Paterson, R. M. C. (1952) J. Chem. Soc. 1718.
- 16. Birkinshaw, J. H. (1965) Biochem. J. 59, 485.
- 17. Malhotra, S. and Misra, K. (1982) Indian J. Chem. 21B, 27.
- 18. Vermes, B., Farkas, L. and Wagner, H. (1980) Phytochemistry
- 19. Hakamori, S. (1964) J. Biochem. 55, 205.